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EFFECT OF TETRAPHENYLBORON UPON THE UPTAKE OF THE LIPOPHILIC CATION DIBENZYLDMETHYLAMMONIUM BY YEAST CELLS

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SUMMARY

The rate of uptake of the lipophilic cation dibenzyltrimethylammonium by yeast cells is increased by tetraphenylboron. However, tetraphenylboron increases also the equilibrium partition of dibenzyltrimethylammonium between cells and medium, probably because a complex between tetraphenylboron and dibenzyltrimethylammonium is trapped inside the cells. Accumulation of dibenzyltrimethylammonium in the presence of tetraphenylboron is not reversed by dinitrophenol, whereas accumulation of the lipophilic cation in the absence of tetraphenylboron appears to be almost completely reversible.

One of the main means of getting information about the membrane potentials of small cells in which it is difficult to measure the membrane potential directly by means of microelectrodes is the determination of the partition coefficient of a lipophilic cation between cells and medium. Frequently the lipophilic quaternary base dibenzyltrimethylammonium, which is able to pass artificial membranes, is applied. This cation has been used with mitochondria [1, 2], bacteria [3, 4] and bacterial vesicles [5, 6]. According to Bakeeva et al. [2] addition of small amounts of tetraphenylboron increases considerably the rate of dibenzyltrimethylammonium uptake by mitochondria and thus causes a decrease in the equilibration time of this cation between mitochondria and medium. This is also found for *Streptococcus faecalis* [3] and for *Escherichia coli* vesicles [6]. The results presented here show that addition of tetraphenylboron may lead to wrong values of the computed membrane potential.

2.2% yeast, *Saccharomyces cerevisiae* Delft 2, is preaerated for 1 night in distilled water in order to deplete the internal substrate of the cells. Then the yeast is washed twice by centrifuging and resuspending the cells in 45 mM Tris/succinate buffer of pH 4.5. The cells are kept anaerobically by bubbling nitrogen through the suspension. ^{14}C -labelled dibenzyltrimethylammonium is added, together with non-labelled dibenzyltrimethylammonium, to the yeast cell suspension to a final concentration of 0.1 mM, whereby the concentration of the yeast is reduced to 2% w/v. When adding tetraphenylboron to the yeast suspension, this is done together with dibenzyltrimethylammonium. Uptake of dibenzyltrimethylammonium is determined

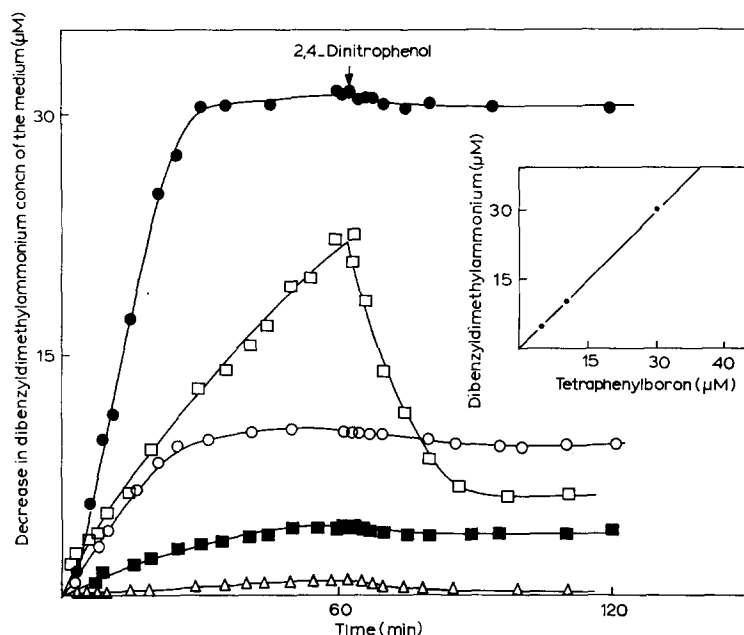


Fig. 1. Effect of varying amounts of tetraphenylboron upon the uptake of 0.1 mM dibenzyltrimethylammonium by non-metabolizing cells, reversal of dibenzyltrimethylammonium uptake induced by adding 1 mM 2,4-dinitrophenol and the relation between the decrease in the concentration of dibenzyltrimethylammonium in the medium and the concentration of tetraphenylboron (inset). On the ordinate is indicated the decrease in concentration of dibenzyltrimethylammonium in the medium. ■, ○, ●, 5, 10 and 30 μ M tetraphenylboron, respectively. □ refers to 20 times the blank (Δ). The arrow indicates the time at which 2,4-dinitrophenol to a final concentration of 1 mM is added.

with the same procedure as that for examining the uptake of radioactive inorganic cation [7], except that washing with acetone is omitted and the radioactivity on the filter papers is measured by means of liquid scintillation according to Borst-Pauwels [8]. 14 C-labelled dibenzyltrimethylammonium is prepared according to Lombardi et al. [5]. Tetraphenylboron is obtained from Sigma chemical company and non-radioactive dibenzyltrimethylammonium from K and K Laboratories.

Fig. 1 shows the kinetics of uptake of dibenzyltrimethylammonium by non-metabolizing cells and the effect of tetraphenylboron upon the kinetics of uptake. Tetraphenylboron increases both the rate of uptake and the equilibrium concentration of dibenzyltrimethylammonium in the cells. The amount of dibenzyltrimethylammonium absorbed by the cell in the presence of tetraphenylboron appears to be almost linearly related to the amount of tetraphenylboron added. As a matter of fact, at an initial concentration of 30 μ M tetraphenylboron, the decrease in dibenzyltrimethylammonium concentration in the medium was also 30 μ M. This indicates that dibenzyltrimethylammonium is trapped in the cells as a stoichiometric 1:1 complex with tetraphenylboron. This view is supported by the fact that the accumulation of dibenzyltrimethylammonium in the cells is no longer reversible in the presence of tetraphenylboron as shown by the fact that 1 mM 2,4-dinitrophenol does not much affect the dibenzyltrimethylammonium concentration in the cells, whereas in the

absence of tetraphenylboron 1 mM 2,4-dinitrophenol leads to an almost quantitative reversal of dibenzyltrimethylammonium uptake.

The uptake of dibenzyltrimethylammonium by metabolizing cells preincubated for 1 h with 3% glucose is also accelerated by tetraphenylboron. The uptake found in the presence of 10 μ M tetraphenylboron is biphasic, consisting of a rapid uptake induced by tetraphenylboron, which is already complete within 1 min, and a slow uptake proceeding at the same rate as the uptake of dibenzyltrimethylammonium occurring in the absence of added tetraphenylboron. The latter uptake is completed after about 30 min, much faster than the uptake by non-metabolizing cells which is completed after about 3 h. Table I shows the effect of 10 μ M tetraphenylboron upon the equilibrium distribution ratios of both metabolizing cells and non-metabolizing cells. The equilibrium distribution ratios represent the quotients of the concentration of dibenzyltrimethylammonium in the cell water (0.44 ml/g of pressed yeast [9]) and the concentration of the cation in the medium.

It is seen that the additional effect upon the distribution ratios exerted by tetraphenylboron does not depend upon the metabolic state of the cells.

There are indications that dibenzyltrimethylammonium forms a complex with tetraphenylboron already in the medium. A solution of 0.1 mM dibenzyltrimethylammonium becomes slightly opalescent on adding tetraphenylboron to a final concentration of 10 μ M or higher. However, it is not probable that the extra amount of dibenzyltrimethylammonium accumulated in the presence of tetraphenylboron consists of adsorption of a complex of tetraphenylboron and dibenzyltrimethylammonium at the outside of the yeast, because the kinetics of the tetraphenylboron dependent uptake of dibenzyltrimethylammonium differs largely for non-metabolizing and for metabolizing cells. The half value times are 15 and ≤ 1 min, respectively, for the two kind of cells.

It may be concluded from the results represented in this article that one should be very careful in using tetraphenylboron as a means for accelerating the rate of dibenzyltrimethylammonium uptake. The apparent partition ratio of dibenzyltrimethylammonium is increased by this compound and too high values for the calculated membrane potential may be found on applying the Nernst equation to the equilibrium distribution data found. There are indications that tetraphenylboron increases the partition ratio of dibenzyltrimethylammonium with other type of cells as well [6, 10]. An increase in dibenzyltrimethylammonium partition ratio is also found with mitochondria when adding an other lipophilic anion, namely phenyl-dicarbaundecaborate, together with dibenzyltrimethylammonium to the mitochondrial suspension [11]. On the other hand, for *Streptococcus faecalis* it is shown that

TABLE I

EQUILIBRIUM DISTRIBUTION RATIOS OF DIBENZYLTRIMETHYLAMMONIUM FOUND IN THE ABSENCE (—) AND IN THE PRESENCE (+) OF 10 μ M TETRAPHENYLBORON

	(—)	(+)
Non-metabolizing cells	1.7	11.5
Metabolizing cells	10.0	20.1

tetraphenylboron does not affect appreciably the dibenzyltrimethylammonium partition ratio [3].

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REFERENCES

- 1 Grinius, L. L., Jasaitis, A. A., Kadziauskas, Y. P., Liberman, E. A., Skulachev, V. P., Topali, V. P., Tsofina, L. M. and Vladimirova, M. A. (1970) *Biochim. Biophys. Acta* 216, 1–12
- 2 Bakeeva, L. E., Grinius, L. L., Jasaitis, A. A., Kuliene, V. V., Levitsky, D. O., Liberman, E. A., Severina, I. I. and Skulachev, V. P. (1970) *Biochim. Biophys. Acta* 216, 13–21
- 3 Harold, F. M. and Papineau, D. (1972) *J. Membrane Biol.* 8, 27–44
- 4 Harold, F. M. and Papineau, D. (1972) *J. Membrane Biol.* 8, 45–62
- 5 Lombardi, F. J., Reeves, J. P. and Kaback, H. R. (1973) *J. Biol. Chem.* 248, 3551–3565
- 6 Hirata, H., Altendorf, K. and Harold, F. M. (1973) *Proc. Natl. Acad. Sci. U.S.* 70, 1804–1808
- 7 Borst-Pauwels, G. W. F. H., Schnetkamp, P. and Van Well, P. (1973) *Biochim. Biophys. Acta* 291, 274–279
- 8 Borst-Pauwels, G. W. F. H. (1968) *FEBS Lett.* 1, 252–254
- 9 Borst-Pauwels, G. W. F. H. and Dobbeltmann, J. (1972) *Acta Bot. Neerl.* 21, 149–154
- 10 De Cespedes, C. and Christensen, H. N. (1974) *Biochim. Biophys. Acta* 339, 139–145
- 11 Griniuvienė, B., Chmieliauskaitė, V. and Grinius, L. (1974) *Biochem. Biophys. Res. Commun.* 56, 206–213